

SYNTHESIS OF THE TETRASACCHARIDE REPEATING-UNIT OF THE O-SPECIFIC POLYSACCHARIDE FROM *Salmonella senftenberg*

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ABSTRACT

The oligosaccharide β -D-Man-(1 \rightarrow 4)- α -L-Rha-(1 \rightarrow 3)-D-Gal-(6 \leftarrow 1)- α -D-Glc, which is the repeating unit of the O-specific polysaccharide chain of the lipopolysaccharide from *Salmonella senftenberg*, was obtained by glycosylation of benzyl 2,4-di-O-benzyl-6-O-(2,3,4-tri-O-benzyl-6-O-p-nitrobenzoyl- α -D-glucopyranosyl)- β -D-galactopyranoside or benzyl 2-O-acetyl-6-O-(2,3,4-tri-O-benzyl-6-O-p-nitrobenzoyl- α -D-glucopyranosyl)- β -D-galactopyranoside with 3-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl- β -D-mannopyranosyl)- β -L-rhamnopyranose 1,2-(methyl orthoacetate) followed by removal of protecting groups.

INTRODUCTION

O-Specific chains of lipopolysaccharides of Gram-negative bacteria are built of repeating oligosaccharide units, the structures of which are unique for each serological type¹. The synthesis of these oligosaccharide units is of interest in studies of the biosynthesis and immunochemistry of microbial polysaccharides, and as a first stage in the chemical synthesis of O-specific polysaccharides.

Recently, we reported on the first synthesis of the repeating unit of the O-specific polysaccharide from *Salmonella anatum* and its analogues^{2,3}, and the β -D-glucopyranosyl analogue of the repeating unit⁴ of the polysaccharide from *S. senftenberg*.

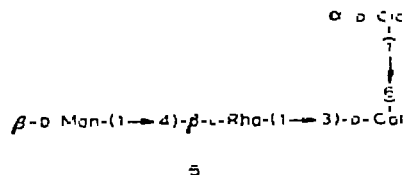
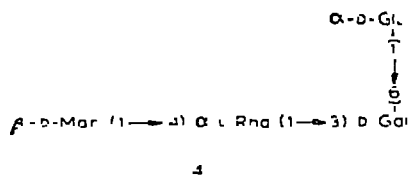
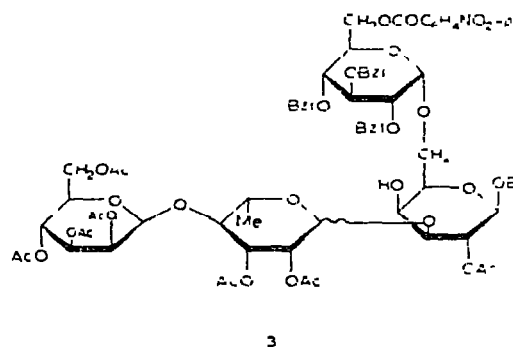
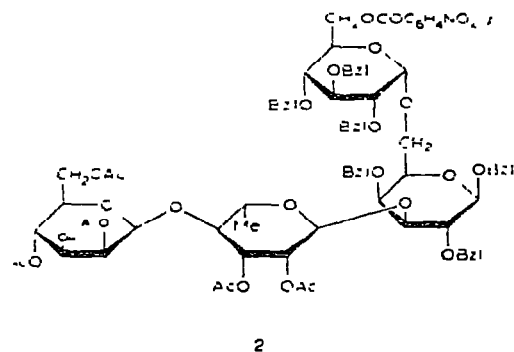
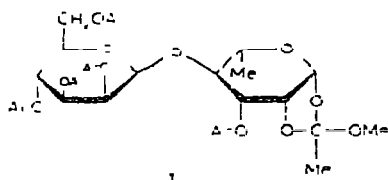
We now describe the synthesis of β -D-Man-(1 \rightarrow 4)- α -L-Rha-(1 \rightarrow 3)-D-Gal-(6 \leftarrow 1)- α -D-Glc, which is the repeating tetrasaccharide unit⁵ of the O-specific polysaccharide of *S. senftenberg* and its α -D-manno analogue α -D-Man-(1 \rightarrow 4)- α -L-Rha-(1 \rightarrow 3)-D-Gal-(6 \leftarrow 1)- α -D-Glc.

RESULTS AND DISCUSSION

A route of synthesis involving consecutive addition of monosaccharide residues from the non-reducing end of the chain could not be developed due to lack of appropriate protecting groups for the synthesis of the α -D-Glc-(1 \rightarrow 6)-D-Gal bond. An

alternative route included the formation of the *O*- α -D-glucopyranosyl-(1 \rightarrow 6)-D-galactopyranose fragment, followed by glycosylation at position 3 of the galactopyranose moiety by a β -D-mannopyranosyl-(1 \rightarrow 4)-L-rhamnopyranose derivative. This scheme is analogous to that used for the synthesis of the glucose analogue of the tetrasaccharide⁴.

Glycosylation of benzyl 2,4-di-*O*-benzyl-6-*O*-(2,3,4-tri-*O*-benzyl-6-*O*-*p*-nitrobenzoyl- α -D-glucopyranosyl)- β -D-galactopyranoside⁴ and benzyl 2-*O*-acetyl-6-*O*-(2,3,4-tri-*O*-benzyl)-6-*O*-*p*-nitrobenzoyl- α -D-glucopyranosyl)- β -D-galactopyranoside⁴ with 3-*O*-acetyl-4-*O*-(2,3,4,6-tetra-*O*-acetyl- β -D-mannopyranosyl)- β -L-rhamnopyranose 1,2-(methyl orthoacetate) (**1**) was carried out with molecular sieve 4 Å as an acceptor of the methanol formed⁶, to yield the respective tetrasaccharide derivatives **2** (27%) and **3** (40%).



The elemental-analysis and p.m.r. data of **2** and **3** were in accord with the expected structures. Deacylation of **2** followed by hydrogenolysis gave the tetrasaccharide β -D-Man-(1 \rightarrow 4)- α -L-Rha-(1 \rightarrow 3)-D-Gal-(6 \leftarrow 1)- α -D-Glc (**4**), which contained rhamnose, mannose, galactose, and glucose residues in the molar ratios 1:1:1:1 as indicated by sugar analysis.

The structure of **2**, expected by the route of synthesis, was proved by identification (g l c - m s) of 3,6-di-*O*-acetyl-1,2,4,5-tetra-*O*-methylgalactitol-1-*d*, which was formed on reduction of **2** with NaBD₄ followed by methylation analysis.

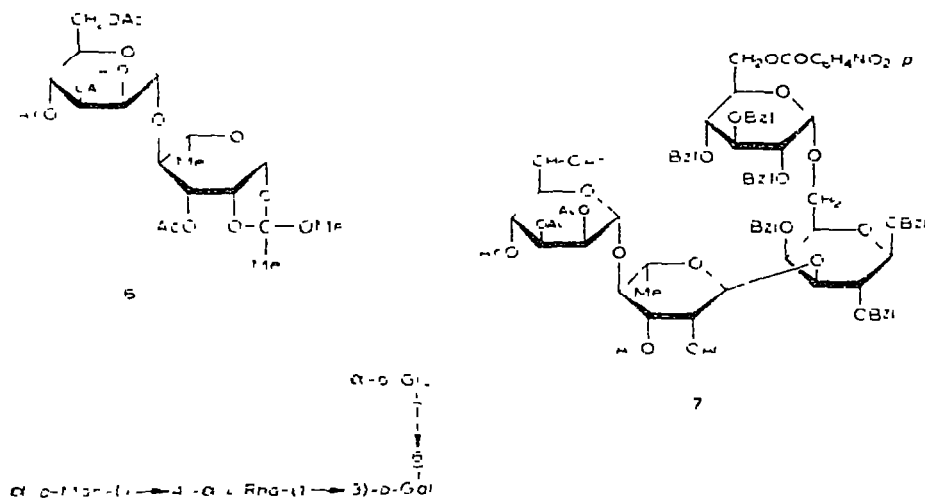
Deacylation of **3** followed by hydrogenolysis afforded a tetrasaccharide which yielded rhamnose, mannose, galactose, and glucose in the molar ratios 1:1:1:1 after acid hydrolysis, and had the same chromatographic mobility as the tetrasaccharide obtained from **2**, but a different optical rotation value. The presence of the rhamnopyranosyl-(1→3)-galactopyranose bond in **3** was confirmed by methylation analysis which gave 1,3,5,6-tetra-*O*-acetyl-2,4-di-*O*-methylgalactitol (identified by g l c - m s).

Ion-exchange chromatography in borate buffer of the tetrasaccharides derived from **2** and **3** showed the former to be homogeneous and the latter to contain two components (**4** and **5**) in the ratio 5:1, the former possessing the same retention time as **4**. The components were isolated by preparative ion-exchange chromatography on Durrum DAX4 resin with borate buffer⁷; on acid hydrolysis, each yielded rhamnose, mannose, galactose, and glucose in the molar ratios 1:1:1:1, but the $[\alpha]_D$ values were **4** +27° (water) and **5** +56° (water).

When the acetates of the tetrasaccharides **4** and **5** were subjected to oxidation with CrO₃ in AcOH, with subsequent hydrolysis and sugar analysis⁸, the rhamnose and mannose residues in the latter were destroyed, but only the mannose residue in the former. Thus, **5** is isomeric with **4** and has a β-L-rhamnopyranosyl bond.

The high selectivity of glycosylation at position 3 in the synthesis of **3** is probably due to the shielding of HO-4 by the glucopyranose residue at C-6, and the formation of a minor proportion of β-glycoside in addition to the α anomer may be attributed to HO-4.

The α-D-manno analogue (**8**) of **4** was prepared via glycosylation of benzyl 2,4-di-*O*-benzyl-6-*O*-(2,3,4-tri-*O*-benzyl-6-*O*-*p*-nitrobenzoyl-α-D-glucopyranosyl)-



β -D-galactopyranoside with 3-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl- α -D-mannopyranosyl)- β -L-rhamnopyranose 1,2-(methyl orthoacetate) (6), followed by removal of protecting groups from the product 7. The structure of 7 and the configuration of the glycosidic bonds were proved in a manner similar to that used for the compounds described above.

EXPERIMENTAL

Melting points were determined with a Kofler apparatus and are uncorrected. P m r spectra were recorded on a Varian DA-60-IL spectrometer with Me_4Si as the internal standard. G l c was carried out with a LHM-8-MD chromatograph and columns of 3% of ECNSS-M on Chromosorb W (1 m) and 5% of SE-30 on Chromaton N-AW (2 m). Mass spectra were obtained with Varian MAT CH-6 and MAT 111 Gnom spectrometers. Optical rotations were determined with a Perkin-Elmer 141 polarimeter. Solutions were concentrated *in vacuo* at 40°. Ion-exchange chromatography of neutral carbohydrates was carried out with a Technicon carbohydrate analyzer on a column (13 \times 0.5 cm) of Durrum DAV4 resin with 0.5M sodium borate buffer (pH 8.54) at 55 and 20 ml/h. The orcinol-sulphuric acid reagent was used to monitor separations. T l c was performed on silica gel "KSK", and p l c on silica gel containing 5% of gypsum. p c was carried out by the ascending method on Filtrak FN 11 paper with (1) chloroform-acetone (95:5), (2) 1-butanol-pyridine-water (6:4:3), (3) chloroform-acetone (9:1). Methylation analysis of oligosaccharide derivatives was performed conventionally.⁸

Syntheses of orthoesters 1 and 6 — The literature procedure⁹ was used, but with lyophilisation of AcOH and HBr from the bromide reaction mixture instead of washing with water³ which gave increased yields (2.5–3 fold) of the orthoesters.

Thus, di-O-acetyl-4-O-(tetra-O-acetyl- β -D-mannopyranosyl)- α -L-rhamnopyranosyl bromide² (1.7 g) gave 1 (1.2 g, 75%), $[\alpha]_{\text{D}}^{20} - 4^\circ$ (c 8, chloroform). P m r data (CDCl_3) δ 3.2 (3 H, O-Me of orthoester), 2.2–1.8 (15 H, 5 Ac), 1.64 (3 H, C-Me of orthoester), 1.22 (d, 3 H, J 5 Hz, C-Me of rhamnose).

Anal. Calc. for $\text{C}_{25}\text{H}_{36}\text{O}_{16}$: C, 50.60, H, 6.09. Found: C, 49.17, H, 6.08.

Di-O-acetyl-4-O-(tetra-O-acetyl- α -D-mannopyranosyl)- α -L-rhamnopyranosyl bromide² (1.2 g) gave 6 (980 mg, 90%), $[\alpha]_{\text{D}}^{20} + 50^\circ$ (c 11, chloroform). P m r data (CDCl_3) δ 3.2 (3 H, O-Me of orthoester), 2.2–1.8 (15 H, 5 Ac), 1.64 (3 H, C-Me of orthoester), 1.22 (d, 3 H, J 5 Hz, C-Me of rhamnose).

Anal. Found: C, 50.20, H, 6.13.

Standard procedure for glycosylation by orthoesters — A solution of 0.3–0.6 mmol of orthoester and 0.2–0.4 mmol of aglycon in dry, freshly distilled CH_3NO_2 (20 ml) was boiled under reflux for 1 h with exclusion of moisture, in a stream of nitrogen and with an interposed molecular sieve (Linde 4 Å). A solution of HgBr_2 (0.1 g) in CH_3NO_2 (10 ml) was added dropwise until reaction was initiated (0.05 mol of HgBr_2 per mol of orthoester). Boiling was continued until reaction was complete.

(~5 h, negative orthoester test, monitoring by t l c) The mixture was then filtered, and concentrated, and the product was isolated by p l c

Synthesis of tetrasaccharide derivatives — (a) Benzyl 2,4-di-O-benzyl-3-O-[2,3-di-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl- β -D-mannopyranosyl)- α -L-rhamnopyranosyl]-6-O-(2,3,4-tri-O-benzyl-6-O-p-nitrobenzoyl- α -D-glucopyranosyl)- β -D-galactopyranoside (2) Benzyl 2,4-di-O-benzyl-6-O-(2,3,4-tri-O-benzyl-6-O-p-nitrobenzoyl- α -D-glucopyranosyl)- β -D-galactopyranoside⁴ {0.2 g, m.p. 125–127°, $[\alpha]_D^{20} + 40^\circ$ (c 2, chloroform)} was glycosylated, as described above, with orthoester 1 (0.2 g) to give 2 (80 mg, 27%), which was isolated by p l c (R_F 0.5, solvent 1) as a syrup, $[\alpha]_D^{20} - 0.075^\circ$ (c 8, chloroform) P m r data ($CDCl_3$): δ 8.0 (4 H, aromatic), 7.2 (30 H, aromatic), 2.0 (18 H, 6 Ac), 1.3 (d, 3 H, J 5 Hz, rhamnose C-Me)

Anal. Calc. for $C_{83}H_{93}NO_{39}$: C, 64.20, H, 5.84 Found: C, 64.23, H, 5.96

(b) Benzyl 2-O-acetyl-3-O-[2,3-di-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl- β -D-mannopyranosyl)- α -L-rhamnopyranosyl]-6-O-(2,3,4-tri-O-benzyl-6-O-p-nitrobenzoyl- α -D-glucopyranosyl)- β -D-galactopyranoside (3) Benzyl 2-O-acetyl-6-O-(2,3,4-tri-O-benzyl-6-O-p-nitrobenzoyl- α -D-glucopyranosyl)- β -D-galactopyranoside⁴ {0.2 g, m.p. 173–175°, $[\alpha]_D^{20} + 22.5^\circ$ (c 2, chloroform)} was glycosylated as described above with orthoester 1 (0.4 g) to give 3 (0.22 g, 40%), which was isolated by p l c (R_F 0.5, solvent 3) as a syrup, $[\alpha]_D^{20} - 9.5^\circ$ (c 2, chloroform) P m r data ($CDCl_3$): δ 8.0 (4 H, aromatic), 7.2 (20 H, aromatic), 2.0 (21 H, 7 Ac), 1.3 (d, 3 H, J 5 Hz, rhamnose C-Me)

Anal. Calc. for $C_{73}H_{82}NO_{30}$: C, 60.03, H, 5.89 Found: C, 60.35, H, 5.80

The acetates of 2,3-di-O-methylrhamnitol, 2,3,4,6-tetra-O-methylmannitol, and 2,4-di-O-methylgalactitol were identified (g l c -m.s.) after deacetylation and methylation analysis of 3

(c) Benzyl 2,4-di-O-benzyl-3-O-[2,3-di-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl- α -D-mannopyranosyl)- α -L-rhamnopyranosyl]-6-O-(2,3,4-tri-O-benzyl-6-O-p-nitrobenzoyl- α -D-glucopyranosyl)- β -D-galactopyranoside (7) Benzyl 2,4-di-O-benzyl-6-O-(2,3,4-tri-O-benzyl-6-O-p-nitrobenzoyl- α -D-glucopyranosyl)- β -D-galactopyranoside⁴ (0.2 g) was glycosylated, as described above, with orthoester 6 (0.2 g) to give 7 (80 mg, 27%) which was isolated by t l c (R_F 0.5, solvent 1) as a syrup, $[\alpha]_D^{20} + 24^\circ$ (c 4, chloroform) P m r data ($CDCl_3$): δ 8.0 (4 H, aromatic), 7.2 (30 H, aromatic), 2.0 (18 H, 6 Ac), 1.3 (d, 3 H, J 5 Hz, rhamnose C-Me)

Anal. Calc. for $C_{83}H_{93}NO_{39}$: C, 64.20, H, 5.84 Found: C, 64.30, H, 6.02

Synthesis of tetrasaccharides — (a) A methanolic solution of 2 (60 mg) was deacetylated conventionally with 0.1 M methanolic sodium methoxide, and then deionized with KU-2 (H^+) resin, filtered, and concentrated. The product was debenzylated over palladium-on-charcoal to give 6-O- α -D-glucopyranosyl-3-O-[4-O-(β -D-mannopyranosyl)- α -L-rhamnopyranosyl]-D-galactose (4, 18 mg, 80%), $R_{LACTOSE}$ 0.34 (p c, solvent 2), $[\alpha]_D^{20} + 27^\circ$ (c 2, water)

(b) By a procedure similar to that described in (a), 7 (60 mg) was converted into 6-O- α -D-glucopyranosyl-3-O-[4-O-(α -D-mannopyranosyl)- α -L-rhamnopyranosyl]-D-galactose (8, 18 mg, 80%), $R_{LACTOSE}$ 0.34 (p c, solvent 2), $[\alpha]_D^{20} + 39^\circ$ (c 2, water)

The tetrasaccharides **4** and **8** were homogeneous on assay with the Technicon carbohydrate analyzer, with retention times of 73 and 125 min, respectively, and gave rhamnose, mannose, galactose, and glucose in the molar ratios 1:1:1:1 after acid hydrolysis (0.5M HCl, 16 h, 100°). The acetates of 2,3-di-*O*-methylrhamnitol, 2,3,4,6-tetra-*O*-methylglucitol, and 1,2,4,5-tetra-*O*-methylgalactitol-1-*d* were identified (g l c - m s) after the reduction of **4** and **8** by NaBD₄ in borate buffer¹⁰ followed by methylation analysis.

(c) By a procedure similar to that described in (a), **3** (0.2 g) was converted into a mixture of **4** and **5** ($R_{\text{LACTOSE}} = 0.34$, p.c., solvent 2), which was fractionated on Durrum DA44 resin with a borate buffer to give 6-*O*- α -D-glucopyranosyl-3-*O*-[4-*O*-(β -D-mannopyranosyl)- β -L-rhamnopyranosyl]-D-galactose (**5**, 10 mg), $[\alpha]_D^{20} + 56^\circ$ (c 1, water), and **4** (50 mg), $[\alpha]_D^{20} + 27^\circ$ (c 5, water), with retention times of 54 and 73 min, respectively.

Acid hydrolysis of **4** and **5** as in (b) gave rhamnose, mannose, galactose, and glucose in the molar ratios 1:1:1:1.

Oxidation of the acetates of tetrasaccharides 4, 5, and 8 — Each tetrasaccharide was conventionally acetylated with acetic anhydride-pyridine and then oxidized¹¹ with CrO₃-AcOH. The products were hydrolysed (2M HCl, 16 h, 100°). By this procedure, the following sugar units were destroyed: mannose in **4**, mannose and rhamnose in **5**, none in **8**.

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